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## Amino Acids as Precursors for N-unsubstituted Fulleropyrrolidine Derivatives.

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Abstract: The formation of azomethine ylides 3 directly from aminoacids and aldehydes and their addition to  $C_{60}$  is reported. The method provides a new route to N-unsubstituted fulleropyrrolidines 4a-4g.

Functionalization of fullerenes continues to be a current focus of research leading to the useful application of fullerene derivatives in biological and materials science. One of the most important methods for C<sub>60</sub> functionalization involves formation of N-alkyl-fulleropyrrolidines by 1,3-dipolar cycloaddition of azomethine ylides to C<sub>60</sub>. This reaction was first reported by Prato<sup>2</sup> and later by some other groups, <sup>3,4</sup> and has served as an excellent and high yield process for fullerene functionalization. <sup>5</sup>

Since N-unsubstituted fulleropyrrolidines provide an entry into further functionalized derivatives by reaction at the nitrogen atom, a pathway to such compounds would be very useful. Prato reported methods for preparing unsubstituted fulleropyrrolidines employing N-trityl protected glycine as a precusor. <sup>2,5b</sup>

Another method, involving the reaction of  $C_{60}$  with an  $\alpha$ -aminoester imine was reported by another group. <sup>4</sup>

We report here the reaction of  $C_{60}$  with azomethine ylides generated directly from aldehydes and amino acids through a decarboxylation route, <sup>6</sup> by which N-unsubstituted fulleropyrrolidines were readily obtained in one step (equation 1).

Azomethine ylides 3a-g were prepared in situ from aldehydes and amino acids as shown in the Table. In a typical procedure, 2 equivalents of amino acid 2, 5 equivalents of aldehyde 1, were mixed in toluene with an equivalent of  $C_{60}$  and heated at reflux for several hours. The rates of reactions depended upon the

reactivities of the ylides, and the reactions could be monitored by TLC. Fulleropyrrolidine products were purified by flash chromatography on silica gel.

Table. Yields and spectroscopic data	<sup>8</sup> for 1,3-dipolar cycloaddition	product 4 (equation 1).
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Entry	Compd	R <sub>1</sub>	R <sub>2</sub>	Yield*	<sup>1</sup> H-NMR (2:1 CS <sub>2</sub> /CDCl <sub>3</sub> , 200 MHz)	
1	4a	Н	CH <sub>3</sub>	20%	4.95 (d, J=12.0Hz, 1H), 4.80 (q, J=6.6Hz, 1H), 4.74 (d	
					, J=12.0Hz, 1H), 2.10 (d, J=6.6Hz, 3H)	
2	4b	Ph	H	35%	7.83-7.79 (m, 2H), 7.46-7.36 (m, 3H), 5.82 (s, 1H),	
					5.12 (d, J=10.7Hz, 1H), 4.90 (d, J=10.7Hz, 1H)	
3	4b	H	Ph	26%	Same as above	
4	4c	H	PhCH <sub>2</sub>	24%	7.50-7.21 (m, 5H), 4.93 (d, J=11.1Hz, 1H), 4.90 (dd,	
1	)	1	1	1	$J_1=3.1Hz$ , $J_2=10.8Hz$ , 1H), 4.65 (d, $J=11.1Hz$ , 1H),	
	ļ				$4.00 \text{ (dd, } J_1=3.1\text{Hz, } J_2=14.1\text{Hz, } 1\text{H), } 3.39 \text{ (dd,}$	
					$J_1=14.1Hz, J_2=11.1Hz, 1H$	
5	4d	Ph	CH <sub>3</sub>	26% trans	trans: 7.83-7.79 (m, 2H), 7.45-7.33 (m, 3H), 5.86 (s,	
ŀ					1H), 5.01 (q, J=6.4Hz, 1H), 2.18 (d, J=6.4Hz, 3H)	
]				23% cis	cis: 7.79-7.76 (m, 2H), 7.45-7.32 (m, 3H), 6.14 (s,	
					1H), 5.39 (q, J=7.0Hz, 1H), 2.25 (d, J=7.0Hz, 3H)	
6	4e	Ph	PhCH <sub>2</sub>	15% trans	trans: 7.86-7.81 (m, 2H), 7.62-7.57 (m, 2H), 7.49-7.32	
					$(m, 6H), 5.71 (s, 1H), 5.04 (dd, J_1=2.7 Hz, J_2=11.1 Hz, $	
	[				1H), $4.05$ (dd, $J_1=2.6$ Hz, $J_2=13.2$ Hz, 1H), $3.53$ (dd,	
i					$J_1=11.1Hz$ , $J_2=13.2Hz$ , 1H), 2.90 (bs, 1H)	
1				20% cis	cis: 7.84-7.79 (m, 2H), 7.56-7.20 (m, 2H), 6.20 (s,	
<b>.</b>	ļ		l		1H), 5.41 (dd, $J_1$ =4.0Hz, $J_2$ =11.5Hz, 1H), 4.05 (dd,	
ĺ					$J_1=11.5$ Hz, $J_2=13.6$ Hz, 1H), 3.72 (dd, $J_1=4.0$ Hz,	
					J <sub>2</sub> =13.6Hz, 1H), 2.93 (bs, 1H)	
7	4f	Ph	Ph	38%	8.06-8.02 (m, 4H), 7.51-7.35 (m, 6H), 6.04 (s, 2H)	
8	4g	PhCH <sub>2</sub>	PhCH <sub>2</sub>	17% <sup>b</sup>	trans: 7.66-7.10 (m, 10H), 4.80 (dd, J <sub>1</sub> =3.1Hz,	
				trans/cis	$J_2=10.5$ Hz, 2H), 3.93 (dd, $J_1=3.1$ Hz, $J_2=13.5$ Hz, 2H),	
				(62/38)	$3.38 \text{ (dd, } J_1=10.5\text{Hz, } J_2=13.5\text{Hz, } 2\text{H})$	
				ļ	cis: 7.66-7.10 (m, 10H), 5.17 (dd, J <sub>1</sub> =3.6Hz,	
					$J_2=10.8$ Hz, 2H), 3.74 (dd, $J_1=3.6$ Hz, $J_2=13.4$ Hz, 2H),	
					3.54 (dd, J <sub>1</sub> =11.1Hz, J <sub>2</sub> =13.5Hz, 2H)	

Isolated yields.

Those reactions using paraformaldehyde gave relatively low yields compared to reactions with other aldehydes. Possibly paraformaldehyde reacts with N-unsubstituted fulleropyrrolidines<sup>6</sup> to produce cross-linked dimers which precipitate from the reaction mixture. Dimeric fullerene derivatives are barely soluble in most organic solvents and characterization of such compounds has proven difficult.<sup>9</sup>

Compounds 4d-4g are 1,3-disubstituted fulleropyrrolidines which may exist as either *cis* or *trans* isomers. That compounds 4d, 4e and 4g exist as *cis/trans* mixtures is shown by <sup>1</sup>H-NMR (table) and HPLC.<sup>10</sup> <sup>1</sup>H-NMR of the *cis*-and *trans*- isomers showed distinctive chemical shifts for the pyrrolidine

b The isomers of compound 4g could not be cleanly separated by flash chromatography. ( $R_f$  value = 0.23 (trans) and 0.19 (cis) on SiO<sub>2</sub>, eluent: 1:1 toluene/hexane).

methine protons. The signals for the cis-isomers always appear further downfield than the corresponding signals for the trans-isomers. (Assignments are based on the stereochemistry expected from steric effects on the known azomethine ylide reaction mechanism, vida infra.) The cis-isomer of 4d shows a methine singlet at 6.14 ppm and a methine quartet at 5.39 ppm, while the corresponding methines from trans-4d appear as singlet at 5.86 ppm and a quartet at 5.01 ppm. Similarly, the pyrrolidine methine signals for cis-4e appears as a singlet at 6.20 ppm and a doublet of doublets at 5.41 ppm. The spectrum of trans-4e shows a singlet at 5.71 ppm, and a doublet of doublets at 5.04 ppm.

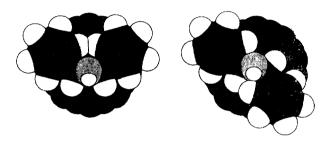


Figure 1. Computer models of cis-4g (a meso compound) and trans-4g (a C<sub>2</sub> symmetric d,l-compound.)

We have previously studied the resolution of chiral C<sub>60</sub> derivatives<sup>11</sup> and thus had an interest in compounds 4f and 4g. The *cis*-isomers of 4f or 4g are *meso* compounds, but the *trans*-isomers are chiral C<sub>2</sub> symmetric racemic compounds (Figure 1). Unfortunately, the reaction leading to compound 4f gave a single isomer as shown by HPLC<sup>10</sup> and <sup>1</sup>H-NMR. This compound was assigned the *cis*-configuration based on its <sup>1</sup>H-NMR spectrum which shows a (downfield) singlet at 6.04 ppm for the pyrrolidine -CH- protons. On the other hand, compound 4g was formed as a 38/62 *cis/trans*-mixture by <sup>1</sup>H-NMR and HPLC. The structure assignments were again based on <sup>1</sup>H-NMR, where the more downfield methine signal (5.17 ppm vs. 4.80 ppm) was assigned to the *cis*-isomer.

The observed differences in chemical yields for cis/trans- isomers could be due to relative stabilities and reactivities of syn- vs. anti-ylides. Based on literature precedent, the trans- isomer must be formed from the anti-ylide and the cis-isomer from the syn-ylide. The syn- and anti-ylides will probably equlibrate before they are trapped by dipolarophiles and it has been suggested that bulky R groups favor formation of the syn-ylides for steric reasons. Our results are consistent with a steric argument since the proportion of cis-compound increases in the series Me->PhCH<sub>2</sub>>Ph.

In conclusion, we report a convenient synthesis of N-unsubstituted fulleropyrrolidines using readily available amino acids as starting materials. Chiral, C<sub>2</sub> symmetric trans-1,3-disubstituted fulleropyrrolidines are suitable for resolution and might be of interest in asymmetric catalysis.

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## References and notes

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